

CAPSULE-BASED BIOIMPEDANCE SENSING FOR INFLAMMATORY BIOMARKER DETECTION IN GASTROINTESTINAL TRACT

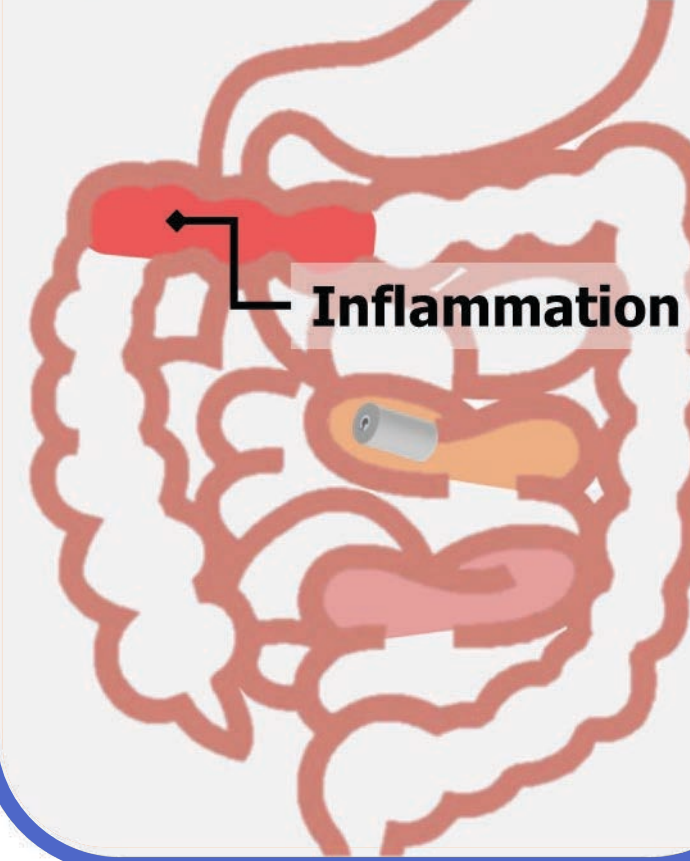
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MEMS Sensors and Actuators Laboratory (MSAL)

MOTIVATION & SIGNIFICANCE

Inflammatory Markers



- Inflammatory Bowel Disease (IBD), comprising Crohn's Disease (CD) and Ulcerative Colitis (UC), is a chronic **inflammatory condition** in the GI tract suspected to be caused by an **unknown** combination of genetic, microbiome, immune, and environmental factors¹
- Pro-inflammatory cytokines IFN- γ and TNF- α **inhibit** intestinal **ion transport channels**, leading to an osmotic pressure imbalance and **leakage** of water and electrolytes through **epithelial tissue**²
- Inflammation linked to an **increase** in **conductive ions** and basal **permeability** of tissue surface

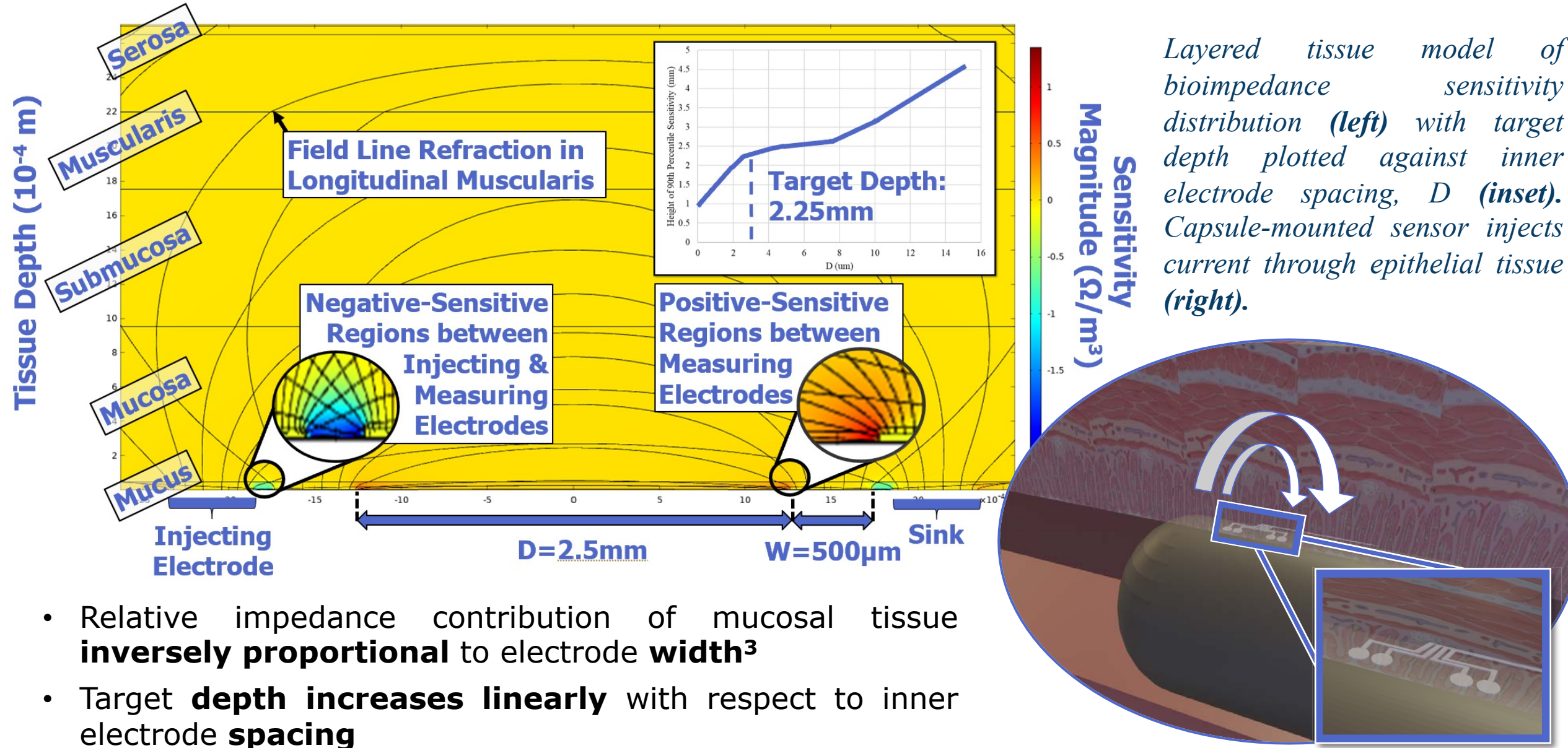
- Few **non-invasive** sensing platforms exist to address **localized tissue inflammation** within **inaccessible** areas of GI tract, such as the small intestine
- Flexible, capsule-mounted sensors to monitor epithelial conductivity **integrate** well with other biomarker sensors for **studying** the **underlying mechanisms** of inflammation

Innovation

- Mounting **bioimpedance sensors** onto **ingestible capsule platforms** enables monitoring of tissue integrity with targeted signal penetration depth
- Breach in mucosal integrity** causes a **decrease** in measured impedance of tissue affected by **inflammation**
- Testing** apparatus to **characterize** bioimpedance sensors traveling along tissue phantom at rate of **peristalsis**

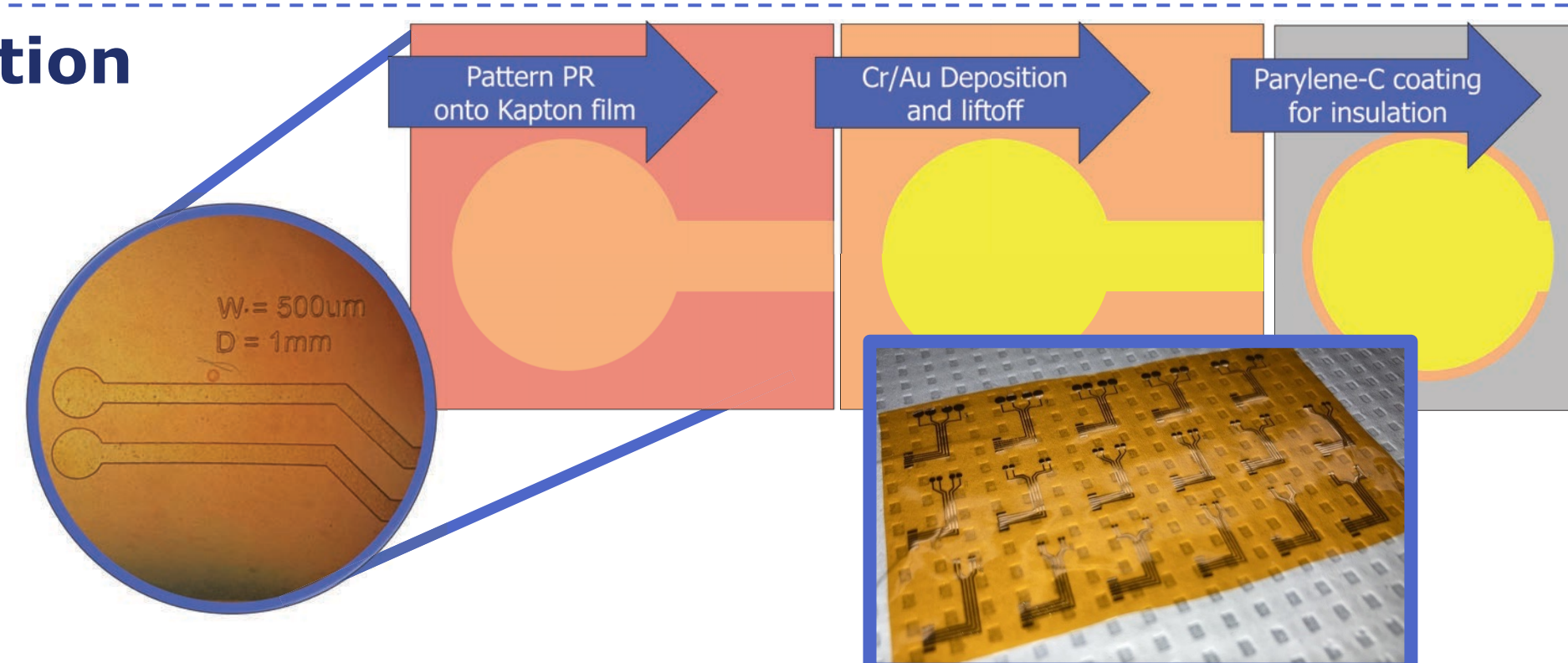
BIOIMPEDANCE SENSOR DESIGN

COMSOL Modeling and Spacing Optimization



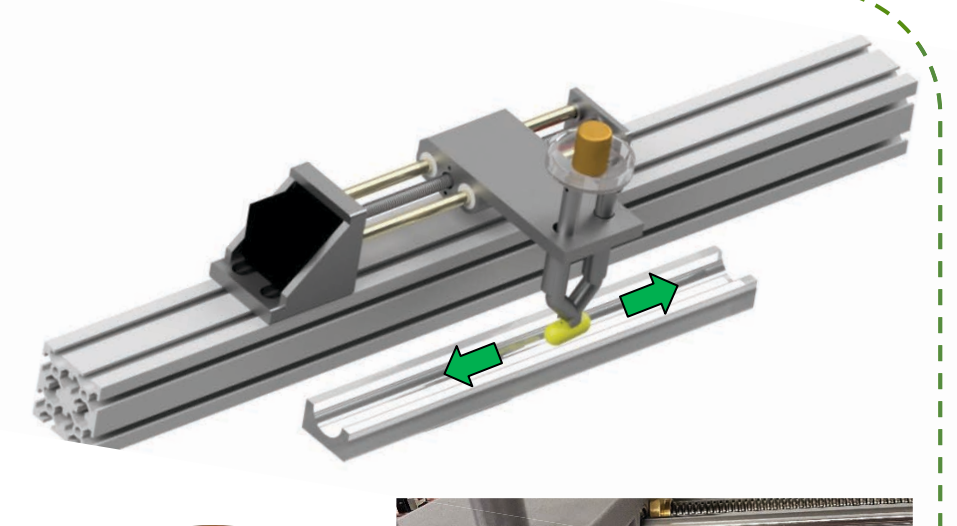
Sensor Fabrication

- Use **photomask** to pattern sensors and deposit **Cr/Au** using electron-beam evaporator
- Parylene-C** will serve as an **insulation layer** to protect traces from shorting



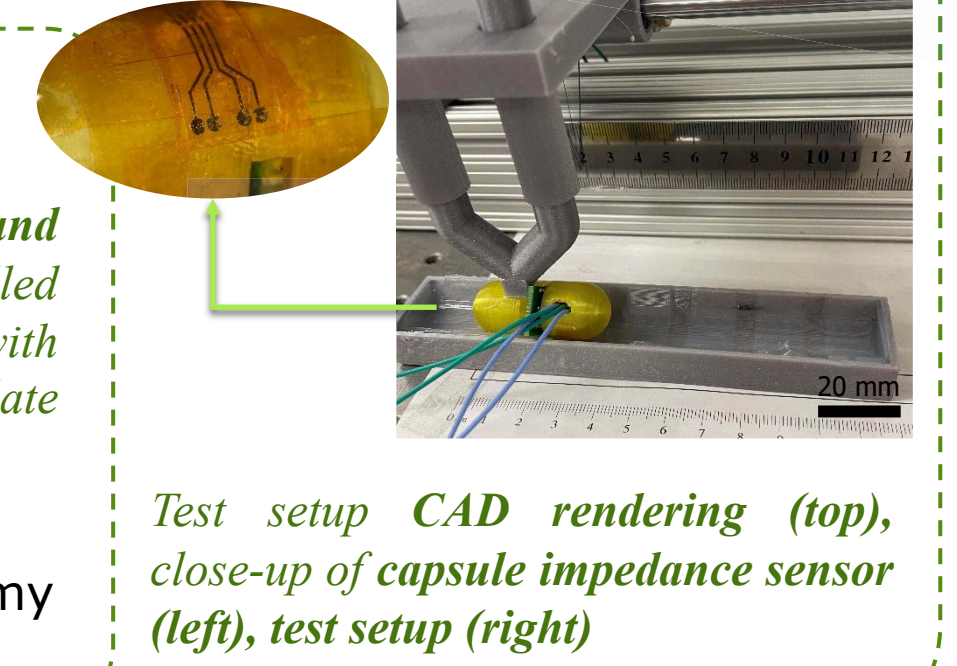
GI SIMULATION SETUP

- Arduino-powered motor moves capsule holder laterally across tissue at a rate of **1.4 cm/min** modeling **peristaltic speeds**⁵
- Sliding member** allows application of **70 gram-force (gf)** force directly on the capsule to model forces experienced in the GI tract⁶
- Allows **testing of drug delivery system**, actuation, and impedance sensor response to various simulated environments



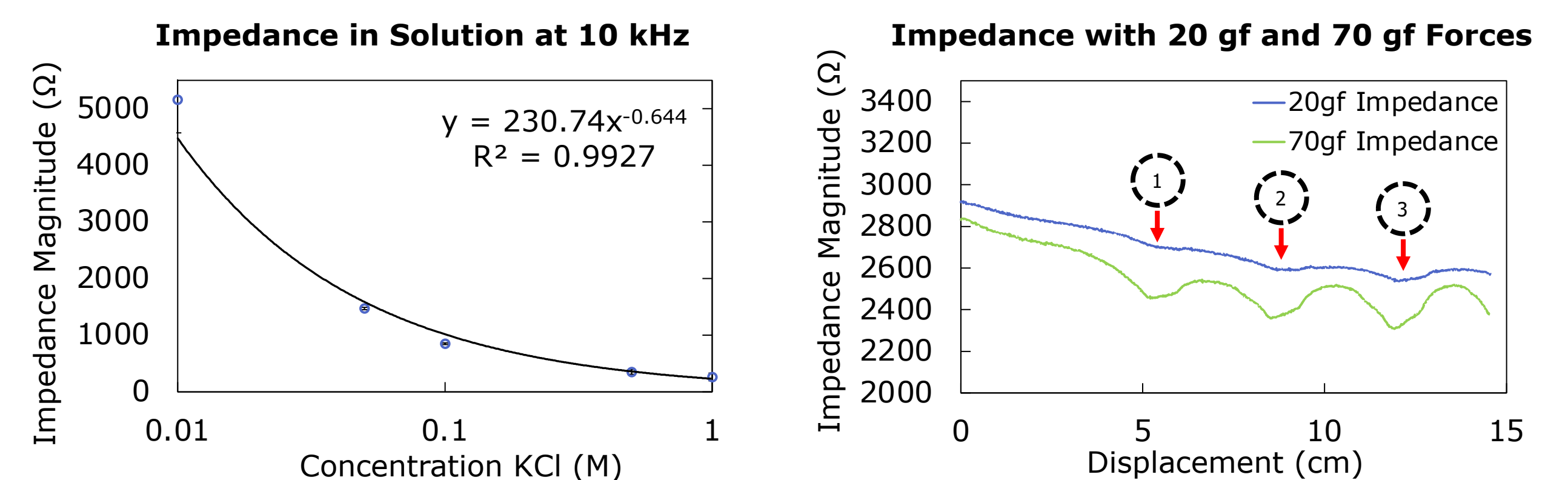
Agar Environment Molding

- Impedance sensors were attached to **3D-printed dummy capsule shells** and displaced using the testing apparatus

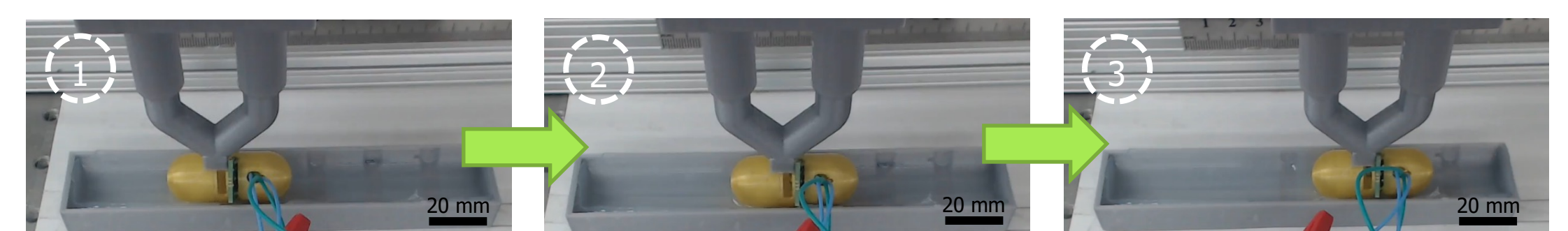


RESULTS

Impedance Sensing Along an Agar Track



Bioimpedance electrodes were characterized (10 kHz) using a Biologic VSP-300 potentiostat at several KCl concentrations (0.01-1 M) ($N=3$, left). The sensor was attached to a capsule shell and actuated across an agar track filled with 1x pH7 PBS. Impedance measurements were recorded while traversing gaps in the mold simulating "non-contact" (right). Forces of 20gf and 70gf, respectively, were exerted on the sensor to evaluate conditions experienced in the GI tract.



Capsule crossing first (left), second (middle), and third (right) agar gaps. Fourth gap not pictured

Significance:

- Quantify **impedance sensor response** to **no contact** with agar in **presence** and **absence of liquid** and **varying applied forces**
- Evaluate impedance sensor **measurement drift** over time
- Assess **impedance sensor response** in various **ion concentrations** to **simulate electrolyte loss** through inflamed mucosal tissue

FUTURE WORK

- Quantify target depth** with respect to electrode width and spacing to validate FEM model
- Use sensors to **characterize permeability** of porcine intestinal **tissue**
- Further investigate the effects of small bowel environmental conditions, such as **contact pressure** and **intestinal viscosity**, on *in vitro* motion artifacts
- Consider effect of **speed** or **acceleration** on impedance sensor results

REFERENCES

- C. Abraham et al., "Inflammatory Bowel Disease," *N. Engl. J. Med.*, 2009;
- D. Magalhaes, J. Cabral, P. Soares-da-Silva, F. Magro, "Role of epithelial ion transports in inflammatory bowel disease," *Am. J. Gastrointest. Liver Physiol.*, 2016;
- P. Kassanos, et al., "A tetrapolar bio-impedance sensing system for gastrointestinal tract monitoring," 2015 IEEE 12th International Conference on Wearable and Implantable Body Sensor Networks (BSN), 2015;
- J.M. Stine et al., "Miniaturized capsule system for hydrogen sulfide detection in the gastrointestinal tract", Hilton Head Workshop 2022: A Solid-State Sensors, Actuators, and Microsystems Workshop 2022;
- M.A. Straker et al., "Region-targeted bilayer coating technology for ingestible devices and systems", Hilton Head Workshop 2022: A Solid-State Sensors, Actuators, and Microsystems Workshop 2022;
- L. Barducci et al., "Fundamentals of the gut for capsule engineers," *Prog. Biomed. Eng.*, 2020.

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